Facile Synthesis of ZnO Flower-Like Micro/nanostructures with Enhanced Antibacterial Activity

Kok Ann Wong¹, Sze Mun Lam^{1*} and Jin Chung Sin²

¹Department of Environmental Engineering, Faculty of Engineering and Green Technology, Universiti Tunku Abdul Rahman, Jalan Universiti, Bandar Barat, 31900 Kampar, Perak, Malaysia

²Department of Petrochemical Engineering, Faculty of Engineering and Green Technology, Universiti Tunku Abdul Rahman, Jalan Universiti, Bandar Barat, 31900 Kampar, Perak, Malaysia

Abstract. Flower-like ZnO micro/nanostructures were synthesized via a simple reflux route without addition of surfactant. The characterization of as-synthesized ZnO sample was performed using X-ray diffraction, field-emission scanning electron microscopy and photoluminescence spectroscopy. The activity of the as-synthesized photocatalyst was evaluated using colony counting method under UV irradiation. The results showed that the as-synthesized ZnO sample exhibited excellent photocatalytic disinfection towards *Escherichia coli*. A possible disinfection mechanism was also postulated based on the findings from microscopic images and radical scavenging experiments.

1 Introduction

Clean water, which is free of pathogenic bacteria and harmful substances is necessary for the human being. In an aqueous environment, the waterborne pathogens including protozoa, fungi, bacteria, prions and viruses and the pathogen-related diseases have caused a considerable risk to public health [1,2]. Thus, it become an essential concern worldwide to secure safe water for the human being. The traditional treatment method that commonly applied is through chemical disinfection. The chemicals such as chlorine and ozone are mostly used for the removal of organic pollutants. Nevertheless, this treatment method often leads to the generation of harmful by-products [3,4].

Recently, the development of light-induced catalytic and antibacterial materials that able to mineralize organic pollutants and inactivate microorganisms has drawn significant attention [4,5]. Irradiation of photocatalyst with light energy results in the formation of reactive oxygen species (ROS), which lead to mineralization of organic pollutants and bacterial inactivation. Among the photocatalysts, zinc oxide (ZnO) has been widely studied because of its large band gap energy (~3.4 eV), large exciton binding energy (~60 meV) and widely applied in solar cells, gas sensors and photocatalysts [6,7]. It is well known that nanostructured photocatalysts can exhibit better properties due to its smaller size, which are

* Corresponding author: lamsm@utar.edu.my

significantly different from those of their bulk counterparts [1]. Until now, various methods such as hydrothermal, electrochemical deposition and sol-gel method have been utilized to produce ZnO nanostructures. The reflux method is more advantageous than other methods in the context of simple, economical, low operating temperature and high yield without the use of any expensive equipment [8]. Moreover, this method can produce nanomaterials with well-defined size and shape without addition of any surfactant or templates [8,9].

In this study, flower-like ZnO micro/nanostructures has been assembled via a simple, productive and surfactant-free reflux route. The characterization of as-synthesized ZnO were then performed by field emission-scanning electron microscopy (FESEM), X-ray diffraction (XRD) and photoluminescence (PL) spectroscopy. The antibacterial activity of as-synthesized ZnO was evaluated against *Escherichia coli* (*E. coli*) under UV illumination. Furthermore, radical scavenging experiment was conducted to determine the main active species that involved in the *E. coli* inactivation. A possible mechanism for the antibacterial activity against *E. coli* over as-synthesized ZnO was also explained.

2 Experimental

In this research, all of the chemicals were in analytical grade and used as-purchased were not further purification. All solutions were prepared with deionized water in this study.

2.1 Construction of flower-like ZnO micro/nanostructures

Zn(NO₃)₂•6H₂O (0.1 M) was first dissolved in 100 mL DI. Then, the solution was added with 100 mL of NaOH (0.1 M) under continuous stirring. Mixture solution pH value was tuned to 12 using extra NaOH and refluxed at 65°C for 8 h. After the solution was cooled down, the precipitates were washed, filtered, dried at 80°C and calcined at 550°C.

2.2 Characterization

The as-synthesized ZnO were investigated by XRD using Philips PW1820 diffractometer with Cu K α radiation at scanning rate of 2° min⁻¹ in the range of 20°-80°. The morphology of the ZnO was analyzed by FESEM JEOL 6701-F. The PL spectroscopy was performed by a Perkin Elmer Lambda S55 spectrofluorometer using a Xe lamp with an excitation wavelength of 325 nm under room temperature.

2.3 Antibacterial activity test

The antibacterial activity test was conducted using colony counting methods. *E. coli* was chosen as the target bacteria. The *E. coli* were cultivated in nutrient broth medium at 37°C. Then, the optical densities of the bacteria were measured at 600 nm using a DR-6000 UV-Vis spectrophotometer. For the photocatalytic inactivation experiment, diluted *E. coli* suspension (10⁵ CFU/mL) was mixed with 400 mL of sterile saline water in a Pyrex reactor. To optimized the bacteria growth, the temperature was set to 37°C. 1 g/L of photocatalyst was then added and continuously stirred for 30 min without light to reach adsorption equilibrium. A Pen-ray UV-C was utilized as a light source for the experiment. At certain time point of UV irradiation, 1 mL of sample was withdrawn out and filtered to remove the suspended solids. Subsequently, the sample was diluted and spread on nutrient agar plates. The colonies on nutrient agar plates were observed after incubated for 24 h at 37°C.

2.4 Detection of reactive oxygen species (ROS)

In order to detect ROS role in photocatalytic inactivation experiment, hydroxyl radicals ($\bullet\text{OH}$), holes (h^+) and superoxide anion ($\text{O}_2^{\bullet-}$) were scavenged by adding 2 mM of isopropanol (IPA), ethylenediaminetetraacetic acid disodium salt (EDTA-2Na) and p-benzoquinone (BQ), respectively. The procedures of radical scavenging test were similar to the antibacterial activity test above.

3 Results and discussion

Fig. 1 presents the FESEM images of the as-synthesized ZnO. As shown in Fig. 1a, the production of ZnO sample was in high yield. It showed that the presence of flower-like structures with varied average size from 800 nm to 2.6 μm . Higher magnification FESEM image in Fig. 1b displayed the flower-like ZnO micro/nanostructures with some petals like protrusion coming out to different direction from the center.

Fig. 1. FESEM images of as-synthesized magnified at different magnifications (a) 5,000x and (b) 10,000x.

Based on FESEM images, a simple growth mechanism for as-synthesized ZnO was proposed. At initial stage, the $\text{Zn}(\text{OH})_2$ was formed when Zn^{2+} cation reacts with OH^- anions. The interior of $\text{Zn}(\text{OH})_2$ was then started to dehydrate and formed large amount of ZnO nuclei. These ZnO nuclei would tend to aggregate together to minimize their surface area under the surface energy driving forces through Ostwald ripening [10]. Subsequently, these aggregates could grow anisotropically along its c-axis and self-assembly into ZnO flower-like micro/nanostructures. Thus, flower-like ZnO micro/nanostructures was successfully assembled via an uncomplicated, low cost and surfactant-free route.

Fig. 2 shows the XRD analysis of the as-synthesized ZnO. The diffraction peaks in XRD result were characterized and well-coordinated with ZnO hexagonal wurtzite structure (JCPDS No. 36-1451) [6,7]. The sharp and narrow peaks implied that the as-synthesized ZnO sample were highly crystalline. In addition, no impurity peaks were detected in the sample, indicating high purity of sample.

Fig. 2. XRD pattern of ZnO flower-like micro/nanostructures.

The PL test was conducted for the ZnO flower-like micro/nanostructures. As shown in Fig. 3, the emission peaks centered at 423 nm, 463 nm, 490 nm and 533 nm were observed. The violet (423 nm) and blue emission (463 nm) were attributed to the defect luminescence or deep level emission from the oxygen vacancies [11,12]. For the green emissions (490 nm and 533 nm), it was due to the oxygen vacancies or deep interstitial oxygen [13]. The result suggested that the ZnO flower-like micro/nanostructures have some oxygen vacancies or deep interstitial oxygen. The oxygen vacancies acted as electron acceptors and temporarily trapped the photogenerated electron (e^-) to decrease the e^-h^+ pair recombination, whereas the deep interstitial oxygen served as trappers for photogenerated h^+ to restrain the recombination of e^-h^+ pair, which improve the photocatalytic performance of photocatalyst [14,15].

Fig. 3. PL spectrum of ZnO flower-like micro/nanostructures at room temperature.

The antibacterial properties of as-synthesized ZnO was evaluated by colony counting method under UV irradiation and the results are presented in Fig. 4. *E. coli* was selected as that target bacteria in this study. *E. coli* can lead to life threatening blood stream infection that related with urinary, gastrointestinal or central nervous systems diseases in human [16]. Control experiments such as photolysis and dark run were conducted prior to the experiment. The results showed that the number of bacteria only decreased slightly under photolysis and dark run. However, under UV irradiation, the ZnO flower-like micro/nanostructures exhibited excellent antibacterial performance towards *E. coli*. Such good performance can be associated to the morphology and oxygen vacancies of ZnO micro/nanostructures. The petal of flower-like structure could penetrate the cell walls of bacteria easily and lead to cell death. Then, the oxygen vacancies could greatly improve the e^-h^+ pair separation which enhanced the photocatalytic performance. Furthermore, the ROS generated could be another reason that terminating the bacterial cells as the ROS could damage the cell membrane and release cellular constituents including lipids and proteins [5,16].

Fig. 4. Photolysis, dark run and photocatalytic antibacterial activity of ZnO flower-like micro/nanostructures towards *E. coli* under UV illumination.

To further interpret photocatalytic disinfection process, radical scavenging tests were conducted to evaluate the main ROS that partook in the photocatalytic disinfection process. From Fig. 5, the antibacterial activity of ZnO flower-like micro/nanostructures only decreased slightly when BQ was added. By adding EDTA-2Na, a noticeably suppression of *E. coli* was observed within 210 min UV irradiation. On the contrary, the antibacterial activity was decreased significantly with the addition of IPA. Thus, these results confirmed that $\bullet OH$ were the main ROS that responsible for the antibacterial activity towards *E. coli*.

Fig. 5. Radical scavenging tests on disinfection properties of as-synthesized ZnO towards *E. coli* under UV illumination.

The morphological changes of *E. coli* before and after the photocatalytic inactivation experiment was observed using FESEM analysis to verify the destruction of *E. coli* cells photocatalyzed by ZnO flower-like micro/nanostructures. From Fig. 5a, well-preserved rod like structure was observed in the untreated *E. coli* cells. On the other hand, the treated *E. coli* cells in Fig. 5b displayed the cells were severely deformed and ruptured, indicating the damage of cell membrane and leakage of cytoplasm. This result further confirmed the antibacterial activity of ZnO flower-like micro/nanostructures toward *E. coli* under UV irradiation.

Fig. 6. FESEM images of *E. coli* (a) untreated and (b) photocatalyzed by as-synthesized ZnO.

According to the experimental results, a possible mechanism was postulated for the antibacterial properties of as-synthesized ZnO and presented in Fig. 7. Upon UV irradiation on as-synthesized ZnO, photogenerated e⁻ was moved from valence band (VB) to conduction

band (CB), leaving h^+ in VB. Photogenerated e^- - h^+ pair was then undergone reduction and oxidation to generate ROS including $\bullet OH$, $O_2^{\bullet -}$ and h^+ . The $\bullet OH$ and $O_2^{\bullet -}$ were the main species the responsible for the photocatalytic disinfection process in this study. Subsequently, these ROS would attack the bacteria cells by degrading the cell membrane and lead to leakage of biomolecules such as carbohydrate, lipids, DNA, nucleic acid and proteins and finally kill the bacteria cell as validated in the FESEM analysis from Fig. 6. Hence, the ZnO flower-like micro/nanostructures was a promising material in photocatalytic disinfection of bacteria.

Fig. 7. Schematic diagram of antibacterial mechanism of as-synthesized ZnO.

4 Conclusions

In this study, flower-like ZnO micro/nanostructures was successfully assembled via an uncomplicated and surfactant-free reflux route. Several analyses including FESEM, XRD and PL were employed to determine the optical and physico-chemical properties of as-synthesized ZnO flower-like micro/nanostructures. The ZnO flower-like micro/nanostructures exhibited excellent antibacterial activity towards *E. coli* under UV irradiation. The radical scavenging tests displayed that $\bullet OH$ were the main ROS that partook in the antibacterial activity towards *E. coli*. Furthermore, the observation of *E. coli* destruction by FESEM further confirmed that the *E. coli* cell membranes were deformed after photocatalyzed by ZnO flower-like micro/nanostructures.

This work was supported by the Universiti Tunku Abdul Rahman (UTARRF/2016-C2/S03 and UTARRF/2017-C1/L02) and Ministry of Higher Education of Malaysia (FRGS/1/2015/TK02/UTAR/02/2 and FRGS/1/2016/TK02/UTAR/02/1).

References

1. M. R. Bindhu, M. Umadevi, M. K. Micheal, M. V. Arasu, N. A. Al-Dhabi, Mater. Lett., **166**, 19 (2016).
2. M. Y. Song, H. D. Jung, J. Jurng, B. C. Kim, Appl. Catal. B: Environ., **148**, 568 (2014).
3. J. Yin, Y. Niu, B. Shao, J. Environ. Sci., **55**, 100 (2017).
4. Y. Zhang, Y. Shao, N. Gao, Y. Gao, W. Chu, S. Li, Y. Wang, S. Xu, Chem. Eng. J., **333**, 85 (2018).
5. Y. Seo, B. E. Yeo, Y. S. Cho, H. Park, C. Kwon, Y. D. Huh, Mater. Lett., **197**, 146 (2017).

6. W. Guo, J. Mater. Sci: Mater. Electron., **27**, 7302 (2016).
7. D. H. Jin, D. Kim, Y. Seo, H. Park, Y. D. Huh, Mater. Lett., **115**, 205 (2014).
8. A.N. Kadam, T. G. Kim, D. S. Shin, K. M. Garadkar, J. Park, J. Alloys Compd., **710**, 102 (2017).
9. K. Mageshwari, R. Sathyamoorthy, J. Park, Powder Technol., **278**, 150 (2015).
10. J. Xie, P. Li, Y. Li, Y. Wang, Y. Wei, Mater. Chem. Phys., **114**, 943 (2009).
11. F. Wang, X. Qin, D. Zhu, Y. Meng, L. Yang, Y. Ming, Mater. Lett., **117**, 131 (2014).
12. B. J. Kwon, J. Y. Kim, S. M. Choi, S. J. An, 2014, Nanotechnology, **25**, 1 (2014).
13. R. K. Biroju, P. K. Giri, S. Dhara, K. Imakita, M. Fujii, ACS Appl. Mater. Interfaces, **6**, 377 (2013).
14. J. Wang, P. Liu, X. Fu, Z. Li, W. Han, X. Wang, Langmuir, **25**, 1218 (2009).
15. X. Sun, J. Wu, Q. Li, Q. Liu, Y. Qi, L. You, Z. Ji, P. He, P. Sheng, J. Ren, W. Zhang, J. Lu, J. Zhang, Appl. Catal. B: Environ., **218**, 80 (2017).
16. S. K. Ray, D. Dhakal, R. P. Pandey, S. W. Lee, Mater. Sci. Eng., C, **78**, 1164 (2017).