Chemical and isotopic characterization of nitrate retained and leached from soil after manure fertilization-by lysimeter experiments

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Abstract. Increase of soil fertilization produces an increase of N exported to the hydrosphere. The amount of nitrate that reaches the aquifers is controlled by processes affecting N-species within the soils. The most relevant processes are nitrification, denitrification, assimilation, mineralization, and immobilization. This work studies the fate of N compounds in soil after manure application in a lysimeter study. To this end the isotopic composition of N and O of dissolved nitrate (δ15N-NO₃⁻ and δ18O-NO₃⁻) was studied coupled with the evolution of N-compounds retained and leached from the soil. Results showed an increase in the δ15N-NO₃⁻ of the leached nitrate towards values similar to the δ15N-NH₄⁺ from the applied manure. The highest δ15N-NO₃⁻ values were measured after 100 days of manure application, and thereafter, values decreased progressively towards the initial δ15N-NO₃⁻ of the soil before manure application.

1 Introduction

Nitrate (NO₃⁻) is one of the most common water pollutants. NO₃⁻ contamination is originated mainly from diffuse (non-point) sources linked to intensive use of synthetic and organic fertilizers and livestock. Increasing use of fertilizers (synthetic and organic) is directly correlated with an increase of nitrate concentration in groundwaters. Previous studies have shown the long lag period between nitrate application to the soil and its discharge to surface and groundwaters, that has been estimated up to several decades [1, 2]. The isotopic characterization of N-compounds is a useful tool to understand processes

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affecting N-compounds in soils. Processes such as nitrification, denitrification, plant assimilation and mineralization-immobilization-turnover (MIT) can modify the isotopic signal of N and O in nitrate, nitrite and ammonium in soils before these compounds are leached to the aquifers [3]. The significance of MIT processes has been reported to affect importantly the $\delta^{18}$O-NO$_3^-$ and limiting its application in nitrate apportionment studies [4]. To gain knowledge on the effect on the isotopic composition of N and O of N-compounds through the processes occurring in the non-saturate zone, a lysimeter experiment was performed to evaluate the isotopic shifts produced in N and O in soils after fertilization with manure. The presence of high organic carbon in manure can favor the production of denitrification reactions. The aim of this work is to characterize isotopically N and O from nitrate, nitrite and ammonium retained and leached from soils after manure application.

2 Methods

2.1 Experimental setup

A lysimeter experiment was set up in a greenhouse at Torre Marimón (IRTA, Caldes de Montbui, Spain). Five lysimeters were constructed. The lysimeters were 48 x 20 cm tubes of polyvinyl chloride, with a perforated lid at the bottom and covered on the top with a 2 mm plastic mesh. The filling material of the lysimeters consisted on a 2 cm layer of rinsed quartz sand, to allow drainage, and a 40 cm layer of a nearby experimental agricultural soil, simulating A and B horizons, equivalent to 13.4 kg of soil. The soil used was obtained from an experimental field that had been fertilized with manure with a low N load (50 kg N/ha·year) during the last 6 years. The lysimeter experiments were installed on March 23$^{rd}$, 2017 and the soil columns were left to stabilize for several days by an initial watering. On April 3$^{rd}$, 2017, fifteen barley seeds (Hordeum vulgare) were sown, and each lysimeter was fertilized with 7.3 g of a thermally dried pig slurry. Five sampling surveys were carried out at relevant stages in relation to plant development and fertilization events (Table 1). An automatic watering was installed on each lysimeter to keep moisture below the maximum water holding capacity (around 20% w/w).

<table>
<thead>
<tr>
<th>Sampling date</th>
<th>Days after setup</th>
<th>Sampling stage</th>
</tr>
</thead>
<tbody>
<tr>
<td>3/4/2017</td>
<td>11</td>
<td>Pre-fertilization</td>
</tr>
<tr>
<td>5/4/2017</td>
<td>13</td>
<td>Post-fertilization</td>
</tr>
<tr>
<td>6/7/2017</td>
<td>76</td>
<td>Developed plant</td>
</tr>
<tr>
<td>5/7/2017</td>
<td>104</td>
<td>Harvest</td>
</tr>
<tr>
<td>4/12/2017</td>
<td>256</td>
<td>Bare soil</td>
</tr>
</tbody>
</table>

2.2 Analytical methods

In each lysimeter, soil, plant and water samples were collected. A 5.5x7 cm core was used to collect topsoil in each lysimeter to prepare soil KCl-extracts followed by a forced leaching. The KCl extracts were immediately prepared by weighting 20 g of soil and adding them to 100 ml of KCl 2M (1:5 v/w), followed by a 30 min-period of shaking in a vertical agitator (120 rpm) (ISO/TS 14256-1: 2003). In parallel, water extracts were prepared using 40 g of soil in a 1:5 ratio (v/w). The KCl and water extracts were centrifuged (5 min at 8000 rpm), filtered in Whatman no. 42 and frozen at -20 °C for later determination of ion
contents. For leachate collection, each lysimeter was placed on a glass tray but raised 1.3 cm to allow drainage. A suitable amount of water was added taking into account the water content at each lysimeter in order to collect around 200 ml of leachate. The leachates were treated and analyzed as described for soil extracts. Plants were harvested at the end of their life cycle in summer. Shoots and leaves were dried at 60°C for 48 h, and finely grounded. N content was obtained by near infrared spectrometry by scanning the grounded samples from 1,100 to 2,500 nm using a NIRSystems 5000 scanning monochromator. NH₄⁺ was measured using the salicylate method, in a Spectronic 20 Genesys 4001/4 spectrophotometer. The NO₃⁻ and NO₂⁻ concentrations were determined by ionic chromatography in an AS4A-SC Dionex anion column in a Dionex DX-100 Ion Chromatograph. The δ¹⁵N and δ¹⁸O of dissolved NO₃⁻ were determined using a modified cadmium reduction method [5, 6]. Simultaneous δ¹⁵N and δ¹⁸O analysis of the N₂O produced was carried out using a Pre-Con (Thermo Scientific) coupled to a Finnigan MAT-253 Isotope Ratio Mass Spectrometer (Thermo Scientific). The δ¹⁵N of dissolved NH₄⁺ was determined using the hypobromite method [7]. The δ¹⁵N of manure applied as fertilizer was determined in a Carbo Erba EA-Finnigan Delta C IRMS.

3 Results and discussion

NO₃⁻ concentrations in K-Cl extracts remained constant until the last sampling (day 245) when its concentration increased drastically (Fig. 1). This increase was likely produced because the last sample was collected 152 days after harvest and during this long period the system was not leached. NO₂⁻ concentrations remained nearly constant along the experiment, ranging from below the detection limit to 0.3 mmol/Kg-soil. Regarding NH₄⁺, the highest values were observed in the first sampling after manure application reaching a maximum of 11.7 mmol/Kg-soil. After this initial peak, values decreased down to 0.3 mmol/Kg-soil and remained constant (Fig. 1). Leachate from the soil showed higher variability between lysimeters with NO₃⁻ concentration between 4.3 and 18.7 mM. In general, NO₃⁻ concentration decreased until day 100, but the last sampling showed higher concentration. NO₂⁻ remained almost constant in all the lysimeters, with values around 0.01, whereas NH₄⁺ showed concentration around 0.03 mM, but also increased sharply in the last sampling (up to 0.5mM). The observed increase of NO₃⁻ and NH₄⁺ could be produced, as in the extracts, because of the long period with no leaching on the lysimeter and the absence of vegetation.

Fig. 2 shows the isotopic composition of manure, irrigation water, and harvested plants, together with the evolution of all the lysimeters leachates and three of the KCl extracts. δ¹⁵N and δ¹⁸O of NO₃⁻ in the soil extracts before manure application ranged between +6.3 ‰ and +4.7 ‰ for δ¹⁵N and +1.7 ‰ and +10.5 ‰ for δ¹⁸O. Regarding leached NO₃⁻, the results of δ¹⁵N ranged from +4.6 ‰ and +8.0 ‰ whereas δ¹⁸O showed values between +2.3 ‰ and +7.6 ‰. Once manure was applied and irrigation started, the isotopic composition of δ¹⁵N increased in both, extracts and leachates. The highest values were measured after 93 days reaching δ¹⁵N values higher than irrigation water but lower than manure. Regarding δ¹⁸O of nitrate a different evolution was observed in some lysimeters. One of them showed higher δ¹⁸O values in the extracts and leachate at day 93 (+12.1 ‰ in soil extracts and +10.1 ‰ in leachate), whereas for the other, values were lower ranging from +4.9 to +7.6 ‰ (Fig. 2). Higher δ¹⁸O could be produced by a slight contribution of denitrification in the first lysimeter. Isotopic results showed that nitrate retained and leached during the first 100 days was mainly influenced by nitrification of manure and nitrate from irrigation producing an increase of δ¹⁵N but maintaining the δ¹⁸O close to the initial values. The slightly higher
δ¹⁵N observed in soil extracts compared with leachate could be related with a higher influence of irrigation water in the leachate. After 245 days, the isotopic data of both the soil extract and the leachate decreased to values close to the initial isotopic composition. This evolution can be explained by a progressive decrease in the effect of fresh manure over the bulk N pool of the soil. As for the δ¹⁸O, the narrow range observed is in agreement with nitrification as the main processes controlling the isotopic composition although a slight contribution of denitrification could not be discarded in some lysimeters.

![Diagram](image1)

**Fig. 1.** Average concentration of NO₃⁻, NO₂⁻ and NH₄⁺ retained in the soil (KCl extracts, left) and leachate (right) along time.

![Diagram](image2)

**Fig. 2.** δ¹⁵N (left) and δ¹⁸O (right) of NO₃⁻ measured in KCl extracts and leachate. Manure, plant and irrigation values are also depicted.

**Conclusions**

Results show the effect of nitrification of N from fresh manure during the first 93 days. Denitrification was not significant, and only in one of the lysimeters the isotopic values suggest a limited effect. Samples collected 245 days after fertilization, showed that the leached nitrate had δ¹⁵N and δ¹⁸O values similar to the initial values of the soil (before fertilization within the experimental period). This fact reflects that the initial N-pool in the soils was very important and masked the influence of the new applied N.

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