

Initial Characterization and Expression Pattern Analysis of Tobacco (*Nicotiana Tabacum*) GMP Synthase Gene

Meiwei Zhao¹, Lei Yang³, Jiacan Wu³, Haijuan Wang³, and Zhengxiong Zhao^{2*}

¹College of Agronomy and Biotechnology, Yunnan Agricultural University, Kunming 650201, China

²College of Resources and Environment, Yunnan Agricultural University, Kunming 650201, China

³Research and Development Center, China Tobacco Yunnan Industrial Co.,Ltd, Kunming 650106, China

Abstract. The complete mRNA sequence of one tobacco (*nicotiana tabacum*) gene—guanosine monophosphate (GMP)synthase, was amplified using the rapid amplification of cDNA ends methods. The full-length tobacco GMP synthase gene mRNA was 2,127bp containing a 1,617 bp open reading frame, which encodes a protein of 538 amino acids. Sequence analysis revealed that the GMP synthase of tobacco shares high homology with the GMP synthase of wood tobacco(99%), *nicotiana attenuata*(99%), *nicotiana tomentosiformis*(99%), potato(92%), *Lycopersicon pennellii*(92%), *lycopersicon esculentum*(92%), *capsicum annuum*(91%), *capsicum chinense*(91%) and *capsicum baccatum*(90%). BLAST analysis within the tobacco high throughout genomic sequences database revealed that this gene has 5 introns and 6 exons. Results also showed that tobacco GMP synthase gene has a closer genetic relationship with the GMP synthase gene of wood tobacco. Tissue expression profile analysis revealed that the tobacco GMP synthase gene was highly expressed in leaf, but moderately expressed in root, flower and stem. Our experiment established the foundation for further research on this tobacco gene.

1 Introduction

GMP synthase or glutamine amidotransferase catalyzes the synthesis of GMP from xantosine monophosphate. This protein is a homodimer, but in some archaea it is a heterodimer composed of a glutamine amidotransferase subunit and a ATP pyrophosphatase subunit. In eucaryotes, bacteria, and some archaea the two catalytic units are encoded by a single gene, producing a two-domain-type GMP, with a GATase domain in the N-terminal half and a ATP-PPase domain in the C-terminal half. The C-terminal domain is specific to the GMP synthases. (<http://www.ebi.ac.uk/interpro/entry/IPR022310>)[1-4].

GMP synthetase is a glutamine amidotransferase from the de novo purine biosynthetic pathway[5]. The amidotransferase family of enzymes utilizes the ammonia derived from the hydrolysis of glutamine for a subsequent chemical reaction catalyzed by the same enzyme. The ammonia intermediate does not dissociate into solution during the chemical transformations[6]. Recent research reported that GMP synthase is essential for viability and infectivity of *Trypanosoma brucei* despite a redundant purine salvage pathway[1].

Although GMP synthase play important roles in GMP biosynthesis of eucaryotes, bacteria and some archaea, until today, the tobacco GMP synthase gene has not been reported yet. In present experiment, we will isolate the complete mRNA sequences of this tobacco

gene, subsequently perform some necessary sequence analysis and tissue expression analysis for this gene. These will establish the primary foundation of understanding this tobacco gene.

2 Material and methods

2.1 Samples collection, RNA extraction and first-strand cDNA synthesis

Tobacco plants (Chinese commercial variety Yunyan 85) were grown in a naturally lit glasshouse with normal irrigation and fertilization. The tissues including leave, stem, root, flower were harvested and immediately frozen in liquid nitrogen and stored at -80°C [7]. Total RNA extraction and first-strand cDNA synthesis for these tissue samples were performed as the methods describe by Li et al.[8].

2.2 5'and 3'-RACE

5'- and 3'-RACE were performed as the instructions of BD SMARTTM RACE cDNA Amplification Kit (BD science, USA). For the tobacco GMP synthase gene, the gene specific primers (GSPs) were designed based on the coding sequence information from potato GMP synthase gene and its highly homologous tobacco EST sequence: EB679319.

5'-RACE GSP:

* Corresponding author: zhaozx0801@163.com

5'- CAATTCACGAACCTCATCCTTGAAT-3'
 3'-RACE GSP:

5'- AAGGCGTAACAGAACCTGAAATGAA -3'.

RACE touchdown PCRs were carried out with 5 cycles of 94°C: 30 sec and 72°C: 3 min, followed by 5 cycles of 94°C: 30 sec, 64°C: 30 sec and 72°C: 3 min, finally with 25 cycles of 94°C: 30 sec, 63°C: 30 sec and 72°C: 3 min to terminate reaction. These RACE PCR products were then cloned into T-vector (TaKaRa, China) and sequenced.

2.3 Quantitative real time PCR (qRT-PCR) for tissue expression profile analysis

qRT-PCR for evaluating the level of mRNA for GMP synthase gene was performed on the ABI Prism 7300 Sequence Detection Systems (Applied Biosystems, Foster City, CA, USA). PCR reactions for each sample were carried out in 25µl reaction volume containing 1µl SYBR Green real-time PCR Master Mix, 100 ng cDNA template and 200 nM each primer. Conditions for real-time PCR were: an initial denaturation at 95 °C for 3 min, 40 cycles of 95 °C for 15 s, 52°C for 15 s (Table 1) and 72°C for 20 s. For each sample, reactions were set up in triplicate to ensure the reproducibility of the results. The gene relative expression levels were quantified relative to the expression of the reference gene, actin (GenBank Accession No. GQ339768) by employing the $2^{-\Delta\Delta Ct}$ value model [7,9].

2.4 Sequence analysis

The cDNA sequence prediction was conducted using GenScan software (<http://genes.mit.edu/GENSCAN.html>).

The protein prediction and analysis were performed using the Conserved Domain Architecture Retrieval Tool of BLAST at the National Center for Biotechnology Information (NCBI) server (<http://www.ncbi.nlm.nih.gov/BLAST>) and the Clustal Omega software (<http://www.ebi.ac.uk/Tools/msa/clustalo/>).

The theoretical isoelectric point (pI) and molecular weight (Mw) of the deduced protein of the tobacco gene was computed using the Compute pI/Mw Tool (http://www.expasy.org/tools/pi_tool.html).

3 Results

3.1 RACE results for tobacco GMP synthase gene

For tobacco GMP synthase gene, through 5'-RACE, one PCR product of 1,265-bp was obtained. The 3'-RACE product was 1,152-bp. These products were then cloned to T-vector and sequenced. Taken together, a 2,127-bp cDNA complete sequence was finally obtained (Figure 1).

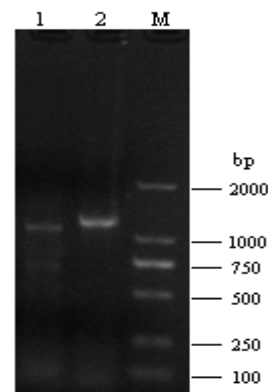


Fig 1. RACE results for tobacco GMP synthase gene. M DL2000 DNA markers; 1, 3'-RACE product for tobacco GMP synthase gene; 2, 5'-RACE product for tobacco GMP synthase gene

3.2 Sequence analysis

These cDNA nucleotide sequence analysis using the BLAST software at NCBI server (<http://www.ncbi.nlm.nih.gov/BLAST>) revealed that this gene was not homologous to any of the known tobacco gene and it was then deposited into the Genbank database (Accession number: KJ011142).

The sequence prediction was carried out using the GenScan software and results showed that the 1,278-bp cDNA sequence represents one single gene which encodes 538 amino acids (Figure 2). The pI of tobacco GMP synthase is 6.51. The molecular weight of this putative protein is 60109.32. BLAST analysis within the tobacco high throughput genomic sequences database revealed that this gene has no intron and is a single exon gene.

Further BLAST analysis of this protein revealed that tobacco GMP synthase has high homology with the GMP synthase of wood tobacco (Accession number: XP_009764602, 99%), *Nicotiana attenuata* (Accession number: XP_019231785, 99%), *Nicotiana tomentosiformis* (Accession number: XP_009600064, 99%), potato (Accession number: XP_006343975, 92%), *Lycopersicon pennellii* (Accession number: XP_015085385, 92%), *Lycopersicon esculentum* (Accession number: XP_004245599, 92%), *Capsicum annuum* (Accession number: XP_016567152, 91%), *Capsicum chinense* (Accession number: PHU28017, 91%) and *Capsicum baccatum* (Accession number: PHT57714, 90%). Its conserved domain was identified as GATase superfamily and GMP_synthase_C (Figure 3).

The 3-D structure of the putative conserved domain also showed that C-terminal of tobacco the GMP synthase contains the GMP_synthase_C domain (Figure 4).

Based on the results of the alignment of different species of GMP synthase proteins, a phylogenetic tree was constructed using the Clustal Omega software, as shown in Figure 5. The phylogenetic tree analysis revealed that the tobacco GMP synthase gene has a closer genetic relationship with that of wood tobacco.

Table 1. qRT-PCR primers for tobacco GMP synthase, actin genes and annealing temperature

| Gene | Primer sequence | Ta/ °C | Length/(bp) |
|--------------|---|--------|-------------|
| GMP synthase | Forward : 5'- CTAATCACCCGCCGAATC-3' Reverse: 5'-GCCACAACCTCAAACCCT -3' | 55 | 410 |
| Actin | Forward :5'-CCATTCTTCGTTTGGACCTT -3' Reverse: 5'- TTCTGGCAACGGAACCT-3' | 56 | 257 |

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ATATAAACCCTAGCCCTCCATTCTCTCTATCTTTCTTCCTCCACCACCACCACCCCTCCCTAGCAATCGAACCTCAAAACACAGCGGAGAAATCAAACTCGTACTTAATCTAGACTACGGTTCTAGTACAC TCAC
CTAATCACCCGCCGAATCCGAAGCCTATCAATTCTCTCACTACCAATTAAACGCCACCTCTCTGGTAGAC TCATAAAAGAACTCCACCCACCTCTGCTATTATCTTCCTGGGTGACCCACAGCGTCCACGGTACGGC
LITRRIIRSLISIFSLITINGTSSLSLDSIKELIDPFRVILISGGPFHVSVAADG
GCACCCTGTTCCACCTGGGTTCATCGAATACCTCGACTCGTGGGATTACCTGTATGCTATGCGGCTCCAGTTGATTGTTAGAAACTGGCGGGTTGTGAAAATGGAGAGAAACATGACTATGGC
APCFPPPGFIEYVESRGIKHVLCICYGLQLLIVLQKLGKGLVQKLGKHEEYF
AGAATGAAATGAGGTGGAAAGAAATGTTGGGGGCTTGGTGGGAAATCGGAAATGGGTATGATAAAGAGCTGGCTTTGGATGAGCCACGGTATGAGGC TGTGAAATTCGGGAAAGGTTTGGGTTGTTGGC GAGG
RMEIEVGEKNVVGGLFFGNTEIGDQVUVVMSHCGDEAVUKLPEGFVVAR
AGTAGTACGGTCTGTTGCTATGGAATCGGAAACGGAGCTTATGCGGCTGCAGTATCACCCGAGCTAAACGCACCTGACGAAAGGATGAGAAC ATTAAGACAC TTTCTGTTGATGTTGGGAT TACA
SSQGCAVA AAIENRRERRFYGLQYHPEVTHSTEGRMRTLRFHFLFDVVCGIT
GCTGGCTGGAAGATGGAAGATCTTCGGAGGAAAGAAATAAAGTATCAAGGCCATGGTTGACCTGAAAGATCACGTGATTTGCTTTATCTGGTGGTGTGATTTCCACTGTCAGCTGAAATGGTACACAGGCT
AGWKMEEDVLEEEIKVIEKMGVCPEDHVICALS GGVDSTVAAKLVHEEA
ATCGGGACAGGCTTTCATGTTTTCGATAATGCTTATAAGGAGAGAGAAAGGCTGATGACCTTTTGGAGAGCCCTCCATTGCTGTGATGATCATAGAAGATTTTCAGC
IGDRLIHCVFVVDNGLLR YKEREERVMDDLFEKRLLHPVTCVDTTEFFLS
AAACTAAAGCGGTAAGAACCTGAAGTGAAGAGGAAATAATGGGAAGGATTCATCAACATATTTGATCTTTTGGCCATGATGTCGAGAAAAAAGTAGGGAAAAACCTTACTTACTAGTCCAAAGAACCTTG
KIKGVVTEPFEKRKRKIIGKEFIWIFDLIFAGHDVEKKEVKGKPTYL VQTIT
TATCTGATGTAATAGACTCTGCTCCCTCCGAGTGGAGAACACATCTCATAACATAGACTCATCATAATGTTGGAGTCTCTTAGGACATGAGCTGAGCTCATAGACCCTGAAACTTCTATTC
YFDVIEESC PPFPSGRTHSHI IKS HVGCEHNVGCALSGGVDSTVAAKLVHEEA
AAGGATGAGGCTCGAATGGGAAAGATTTGGATATATCGAGACTTCTTAAAGCCACCGCTTCCGCGCCGGCTGTACGAACTCAGGATGATGTCACAGCGGGAAATTCCTGGATATTCCTG
KDEVRHELGLKILDISEDFLKRHPFPGLAVRIPFDVTRASLDILR
CAGGTTGATGATCTTCAATTCATCAGAGATGCTAAATCTATGATGAAATATGGCAGCTTTTGGCTTCTTACAGTGAAACTGTTGGAGTACAGGAGACCAAGAACACATCCCATGCTGTGCA
QVDETIQSIRDAKTYDEIWAFAVFLPVKTVG VGGDRTHSHAV
CITAGAGCAGTACAGCTAAGATGGAACTCGACTCTTGAATTCAGCTTCTGACGACTATCAAGAAAGATCTGCAATAGCTCTTCAGTCTTCAATAGATATACATAAG
LRAVTSQDGM TADWY YFD FKLDDVSRKICHSVURGVNRVLLDITTSK
CCTCCATCAACATCGAGTGGCAATATTGTTTAAAGAAATGCTATATTTGGTGCACCAAGCTAGGATCTTTTGTGATTTTGGTGCATAACAAAAGGAAAGAAATCATGATAGAAATTAGGCTCTTTTGTATGTCG
E...E...S...T...I...E...W...E...
CTAGAACCTGCTTTGGGCTAATTATGTCAGTCTTCCGACAAATTTTGTACCGGGGAAATGCAATTTGATGGATGCTCCGCGCATGAGAGAGGAGGACGCTTTCAGAGCGGAAAGGCCATGGGGAGATCCCTCTGATC
CATGGATCTCCGATCGGAAACCGTATCCAGCTCCCTGGCTAGTTTGGCTCTTTGGACCTTTTCAAACTAGGGAAC TGAACATCTTAGTAGCTAAAGGAGGGGAAATCAACCAGACCCCTTAGCTAGCGCCGAGCG
AGAGCGGATTTGGGCTTTCAGAAAAAAGGAAAAAAAAAAAAAAAAAAAAAAAA
    
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Fig 2. The complete mRNA of tobacco GMP synthase gene and its encoding amino acids *indicates the stop codon

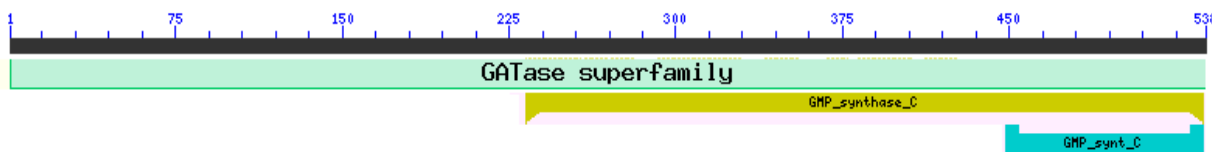


Fig 3. The putative GATase superfamily domain and GMP_synthase_C domain of the protein encoded by tobacco GMP synthase gene

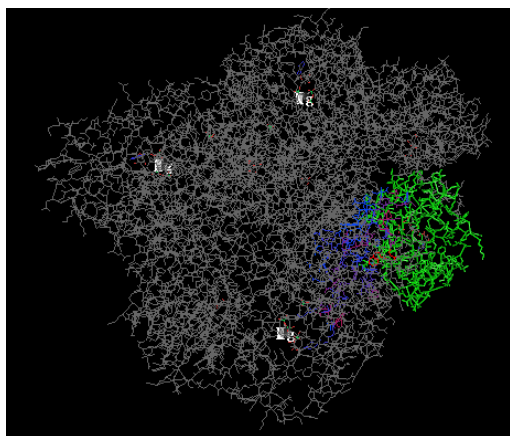


Fig 4. The 3-D structural evidence of the putative conserved GMP_synthase_C domain of tobacco GMP synthase protein

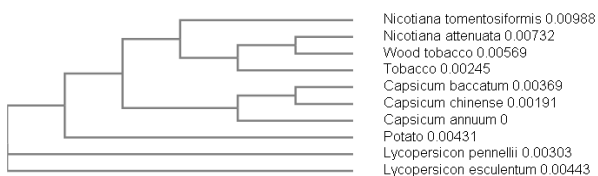


Fig 5. The phylogenetic tree for six kinds of GMP synthase gene

To obtain the genomic DNA of common tobacco GMP synthase gene, the publicly available common tobacco whole genome shotgun sequence database was screened using the coding sequences of common tobacco GMP synthase gene as seeds by BLAST analysis. A whole genome shotgun sequence (GenBank accession no. NW_015883288.1) was found to encompass the entire GMP synthase gene (8,071bp-12,578bp). The common tobacco GMP synthase gene is 4,508-bp in length and consists of 6 exons and 5 introns. All exon–intron splice junction sequences conform to the GT–AG rule (Figure 6).

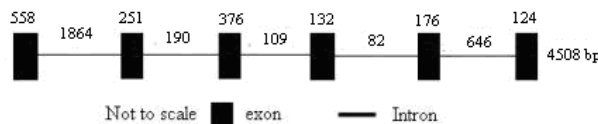


Fig 6. The genomic sequence organizations representing the coding regions of the common tobacco GMP synthase gene

3.3 Tissue expression profile

Tissue expression profile analysis was carried out and results revealed that the tobacco GMP synthase gene was highly expressed in leaf, but moderately expressed in root, stem and flower (Figure 7).

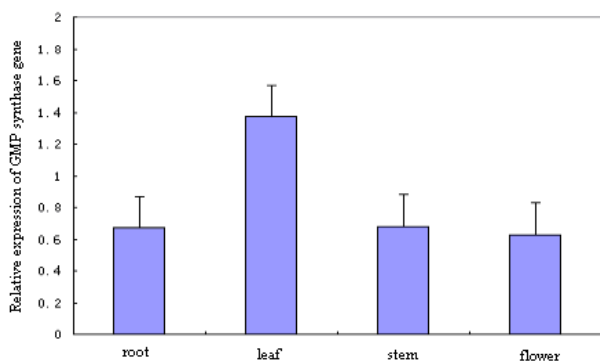


Fig 7. Expression analysis of GMP synthase gene mRNA in various tissues

4 Conclusions

Protein comparison analysis revealed that tobacco GMP synthase shares high homology with several GMP synthetases. GMP synthetase is a bifunctional two-domain enzyme. a GATase domain in the N-terminal half and a ATP-PPase domain in the C-terminal half which is specific to the GMP synthetases[1-4]. In our results, the putative conserved domain analysis showed that of tobacco GMP synthase contains the GATase superfamily and GMP_synthase_C conserved domains. These imply that our study results are reliable.

The phylogenetic tree analysis revealed that the tobacco GMP synthase gene has a closer genetic relationship with that of wood tobacco. This implied that we can use wood tobacco as model organism to study the tobacco GMP synthase gene or use tobacco as model organism to study the wood tobacco GMP synthase gene. From the tissue distribution analysis in our experiment it can be seen that GMP synthase gene was obviously differentially expressed in some tissues. The tobacco GMP synthase gene was highly expressed in leaf, but moderately expressed in root, stem and flower. For GMP synthase catalyzes the synthesis of GMP from xantose monophosphate[1-4]. The suitable explanation for differential expression under current conditions is that the GMP biosynthesis was high in leaf, but moderate in root, stem and flower.

In conclusion, we first isolated the tobacco GMP synthase gene and performed necessary sequence analysis and tissue expression analysis. This established the primary foundation for further research on this tobacco gene.

Acknowledgement

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