

Volatile fatty acids concentration and efficiency of microbial protein synthesis of concentrate cassava peel diet with different levels of protein sources

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Abstract. This study was conducted to evaluate the effects of different levels of protein sources in a concentrate cassava peel diet on volatile fatty acid (VFA) concentration and efficiency of microbial protein synthesis (EMPS) by *in vitro* method. The five diet treatments in this study consisted of cassava meal (CM), cassava peel (CP), cassava leaves (CL) and moringa leaves (ML) with the proportion applied were T1=CM70% + CP20% + CL5% + ML5%; T2=CM60% + CP20% + CL10% + ML10%; T3=CM50% + CP20% + CL15% + ML15%; T4=CM40% + CP20% + CL20% + ML20%; T5=CM30% + CP20% + CL25% + ML25%. The experiment used a randomized block design with five treatments and three replications. The results showed that increasing the level of protein sources in the ration tended to increase the value of total VFA, acetate, propionate, CO₂, and CH₄ in which T3 had the highest values. However, the effect of treatments were significant in EMPS value (P<0.01). It was concluded that increasing levels of cassava and moringa leaves in the concentrate cassava peel diet tended to increase the value of total VFA, acetate, propionate, butyrate, CO₂, and CH₄ which led to a significant increase in EMPS values.

1 Introduction

Carbohydrates are fermented by rumen microbes to produce volatile fatty acids (VFA) which consist of 60-70% acetic acid, 15-20% propionic acid, and 10-15% butyric acid [1]. The concentration of VFA is influenced by the level of feed ingredient fermentability in the rumen [2]. Apart from their function as a source of energy for the host, VFAs also serve as a source of carbon skeletons as well as ATP for microbial protein synthesis [3]. The microbial protein produced in the rumen and flows into the abomasum provides the majority of protein for further digestion by HCl-pepsin and trypsin to form amino acids in the small intestine for absorption (50-80%). Bach *et al.* [4] concluded that rumen ammonia is the main precursor,

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and its efficiency of conversion into microbial protein is determined by the type and amount of feed consumed by ruminants. Ramaiyulis *et al.* [5] stated that rumen microbial growth, largely determined by the availability of essential nutrients such as soluble carbohydrates and protein.

Cassava (*Manihot utilissima* Crantz) is a major plant in Indonesia with 19.3 million tons production in 2018 [6]. This perennial plant contains high energy in the root [7] and a high protein in the leaves [8]. Cassava meal which formed from the drying and grinding process of cassava tubers contains 87.3% dry matter (DM), 2.13% crude protein (CP), and 3.2% crude fiber (CF) [9]. Lukuyu *et al.* [10] mentioned that cassava meal provides 96.2% organic matter (OM) and non-fiber carbohydrates (NFC) 82%. Cassava peels contained 7.1% OM, 7.1% CP, 2% extract ether (EE), and 66.4% NFC. Hang *et al.* [11] mentioned that cassava leaves are rich in protein with 29.4% DM and 22.1% CP.

Retnaningrum *et al.* [12] reported that the use of cassava meal as an energy source to supplement maize stover basal diet had the highest digestibility and daily weight gain of crossbred Limousine. [13] mentioned that cassava peel can replace corn as an energy source in ration rated from ruminal parameters on small ruminants. Mulyati *et al.* [14] stated that cassava and moringa leaves potentially as a supplement on low-quality roughages to support rumen microbial growth and activity due to high protein content. This study aimed to evaluate the effects of different levels of protein sources in a concentrate cassava peel diet on VFA concentration and efficiency of microbial protein synthesis (EMPS) by *in vitro* method.

2 Materials and methods

2.1 Location and time

This research was conducted in the Nutrition and Animal Feed Laboratory Faculty of Animal Science Brawijaya University in November.

2.2 Materials

Materials used were set of tools for volatile fatty acids (VFA) and efficiency of microbial protein synthesis (EMPS) analysis and feedstuffs consisted of cassava meal (CM), cassava peel (CP), cassava leaves (CL) obtained from Ngajum, Malang, and moringa leaves (ML) took from Kambingan Village, Tumpang, Malang.

2.3 Method

This study used a Randomized Block Design (RBD) with five treatments and three replication. The treatments applied were T1=CM70% + CP20% + CL5% + ML5%; T2=CM60% + CP20% + CL10% + ML10%; T3=CM50% + CP20% + CL15% + ML15%; T4=CM40% + CP20% + CL20% + ML20%; T5=CM30% + CP20% + CL25% + ML25%. Table 1 showed the chemical composition of feedstuffs and treatments.

The *in vitro* method used Makkar *et al.* [15] protocol, in which after the incubation, the residue were centrifuged to separate sediment and supernatant parts. The separated sediment was dried and weighed for measuring fermented organic matter (FOM) value. The separated supernatant was used to determine the VFA total, acetate (C₂), propionate (C₃), and butyrate (C₄) by gas chromatographic techniques. The result of VFA concentration also used to determine CO₂ and CH₄ by Van Soest [16] equation. For determining of EMPS value, the residues were refluxed using NDS solution and calculated with Bach *et al.* [4] equation.

Table 1. Chemical composition of feedstuffs and treatments

Feed	Chemical Composition (%)							
	DM	Ash*	CP*	EE*	CF*	NFE*	ADF*	NDF*
Cassava Meal (CM)	84.41	2.63	2.86	0.84	2.90	90.77	2.18	8.96
Cassava Peel (CP)	24.32	7.51	9.18	1.72	9.88	71.72	9.36	28.36
Cassava Leaves (CL)	25.73	9.87	25.48	9.72	18.46	36.47	47.15	46.50
Moringa Leaves (ML)	24.23	12.97	27.38	6.78	10.35	42.51	13.39	17.51
T1	90.62	4.26	9.10	1.22	4.76	80.66	7.34	16.03
T2	91.47	5.03	11.50	1.69	5.66	76.12	8.50	17.50
T3	91.87	5.98	12.56	2.35	6.82	72.29	10.19	19.03
T4	92.29	6.94	14.46	3.16	7.56	67.88	11.68	22.57
T5	90.88	7.87	19.28	4.65	8.57	59.62	13.09	33.14

*) Based on 100% of DM [17]

2.4 Statistical analysis

The data obtained were analyzed by analysis of variance (ANOVA) and followed by Duncan’s Multiple Range Test if the treatments gave a significant effect on the variables measured according to Sudarwati *et al.* [18].

3 Results and discussion

Table 2 presents the result of VFA total, acetate, propionate, butyrate, CO₂, CH₄, and EMPS by treatments. The results showed the treatment did not significantly affect the value of all parameters (P>0.05), except EMPS which showed a significant by treatments (P<0.01). Treatment T3 achieved the highest value of total VFA (148.6 mMol/l), whilst the lowest total VFA value was recorded in T1 (97.4 mMol/l). Similar trends were observed in acetate, propionate, butyrate, CO₂, and CH₄ values. Increased proportions of cassava and moringa leaves in the ration has led to EMPS value being highest for treatment T5 (27.28 g N/kg FOM), whilst T2 had the lowest value of EMPS (12.10 g N/kg FOM). This evidence indicates the potential of cassava and moringa leaves as protein sources to supplement cassava peel and cassava meal as carbohydrate sources.

The VFA total in this study was high because the concentration of VFA was generally from 70-150 mMol [19] and this may be due to a high fermentation rate of cassava meal and peels in the rumen. The high starch diet in ration increases acetate and propionate value [20] and increases the amyolytic microbial population [19]. Suharlina *et al.* and Gumilar *et al.* [21, 22] stated that fibrous feed becomes acetic in the rumen which probably caused acetate and propionate to be higher. In contrast, T3 has a higher concentration of VFA than T4 and T5 which have higher fiber. It also means that the high protein content in the ration without being followed by an adequate energy source cannot be utilized properly by microbes. Retnaningrum *et al.* [12] reported that the VFA increased with the addition of cassava meal in the ration, but at the 70% proportion of cassava meal, drastically decreased total VFA,

acetate, propionate, and butyrate due to inadequate proportion of protein source. Santos *et al.* [13] reported that cassava peel tended to decrease propionate and increase the acetate concentration in replacement of corn.

Table 2. Total VFA, acetate, propionate, butyrate, CO₂, CH₄ and EMPS of treatments

Parameters	Treatments					SEM
	T1	T2	T3	T4	T5	
Total VFA (mMol/l)	97.40	133.10	148.63	136.13	137.60	8.298
Acetate (mMol/l)	61.67	86.43	95.63	87.40	85.30	5.538
Propionate (mMol/l)	21.63	28.0	30.53	29.90	30.67	2.074
Butyrate (mMol/l)	14.10	18.67	22.47	18.83	21.63	1.516
CO ₂ (Mol)	58.86	58.82	60.21	57.90	60.0	0.688
CH ₄ (Mol)	32.63	34.19	34.77	33.74	33.25	0.841
EMPS* (g N/kg FOM)	12.71 ^a	12.10 ^a	17.37 ^{ab}	20.11 ^{ab}	27.28 ^b	1.910

Different superscript in the same row showed the significant effect at P<0.01 (*)

EMPS= Efficiency of Microbial Protein Synthesis

FOM= Fermented Organic Matter

Apart from VFA levels, CH₄ can be assessed for feed efficiency parameters. In this study, the CH₄ trend was similar to the VFA value which the highest value was T3 (34.77 Mol) and the lowest in T1 (32.63 Mol). McDonald *et al.* [1] mentioned that the organic matter degradation by rumen microbes will produce VFA, CO₂, CH₄, and other gasses. The higher VFA formed higher the gasses produced. Muchlas *et al.* [23] stated that the higher CH₄ production illustrates more energy wasted, which indicates the more inefficient the feed was.

Karsi and Russel [24] mentioned that EMPS values generally range from 10-70 g N/kg FOM and the optimal value for EMPS above 30-40g N/kg FOM, thus the EMPS value in this study is low. It may be caused by the low fiber diet and a high concentrate that caused the low rumen pH as stated by Muchlas *et al.* [23]. The low EMPS in this study may be causing a high value of NH₃, in which according to Bach *et al.* [4], the available energy and amino acid (AA) will be used directly for microbial protein synthesis, while limited energy will cause amino acid (AA) to be deaminated and the carbon skeleton fermented into VFA. The excess of AA will be removed from the cytoplasm into ammonia because some rumen microbes lack of mechanism for emitting AA.

4 Conclusion

In conclusion, increasing levels of cassava and moringa leaves in the concentrate cassava peel diet tended to increase the value of total VFA, acetate, propionate, butyrate, CO₂, and CH₄ which led to a significant increase on EMPS values.

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References

1. P. Mc Donald, R.A. Edwards, J.F.D. Greenhalgh, C.A. Morgan, L.A. Sinclair, and R.G. Wilkinson, *Animal Nutrition 7th Edition* (Benjamin Cummings, 2011)
2. C.J.B. Sendow, C.T. Noviandi, and R. Utomo, *Buletin Anim. Sci.* **41** (2017)
3. E.R. Orskov, *Protein Nutrition in Ruminants, 2nd Edition* (Academic Press, 1992)
4. A. Bach, S. Calsamiglia, and M.D. Stern, *J. Dairy Sci.* **88**, 9-21 (2005)
5. Ramaiyulis, R.W.S. Ningrat, M. Zain, and L. Warly, *Pak. J. Nutr.* **18** (2019)
6. A. Amir, B.P. Purwanto, Nahrowi, A. Atabany, Salundik, and A. Yani, *LRRD* **33** (2021)
7. P. Sumadong, A. Cherdong, S. So, and M. Wanapat, *BMC. Vet. Res.* **17** (2021)
8. S. Suharti, H. Oktafiani, A. Sudarman, M. Baik, and K. G. Wiryawan, *AOAS.* **66** (2021)
9. S. Retnaningrum, Kusmartono, Mashudi, K.J. Harper, and D. Poppi, *IOP Conf. Ser. Earth Environ. Sci.* **478** (2020)
10. B. Lukuyu, I. Okike, A. Duncan, M. Beveridge, and M. Blummel, *Use of Cassava in Livestock and Aquaculture Feeding Programs* (ILRI Editorial and Publishing Services, Addis Ababa, Ethiopia, 2014)
11. L.T.T. Hang, T.R. Preston, N. Xuan Ba, and D. Van Dung, *LRRD.* **31** (2019)
12. S. Retnaningrum, Kusmartono, Mashudi, K.J. Harper, and D.P. Poppi, *Animal*, **15**, 1-7 (2021)
13. V.L.F. Santos, M.A. Ferreira, M.C.B. Siqueira, T.T.B. Melo, J.L. Silva, I.B. Andrade, A.A. Soares, and C.T.F. Costa, *Small Rumin. Res.* **113**, 88-92 (2015)
14. Mulyati, Kusmartono, Hartutik, and Rusdi, *LRRD*, **27** (2015)
15. H.P.S. Makkar, M. Blümmel, and K. Becker, *British J. Nutri.* **73**, 897-913 (1995)
16. J. P. Van Soest, *Nutrition Ecology of Ruminant, 2nd Edition* (Cornell University Press, Ithaca, 1994)
17. A.D. Saputra, Kusmartono, Mashudi, P.H. Ndaru, *IOP Conf. Ser. Earth Environ. Sci.* **888** (2021)
18. H. Sudarwati, M.H. Natsir, and V.M.A. Nurgiartiningsih, *Statistika dan Rancangan Percobaan dalam Bidang Peternakan* (Universitas Brawijaya Press, Malang, Indonesia, 2019)
19. A.D. Putridinanti, C.T. Noviandi, Gunawan, A. Agus, K. Harper, and D. Poppi, *IOP Conf. Ser. Earth Environ. Sci.* **387** (2019)
20. B. Hatew, S.C. Podesta, H. Van Laar, W. F. Pellikaan, J.L. Ellis, J. Dijkstra, and A. Bannink, *J. Dairy. Sci.*, **98**, 486-499 (2015)
21. Suharlina, D.A. Astuti, Nahrowi, A. Jayanegara, and L. Abdullah, *JITAA*, **41** (2016)
22. D.A.K.W. Gumilar, E. Rianto, and M. Arifin, *IOP Conf. Ser. Earth Environ. Sci.* **119** (2018)
23. M. Muchlas, Kusmartono, and Marjuki, *Jurnal Ilmu-Ilmu Peternakan*, **24**, 8-19 (2014)
24. M.A. Karsli and J.R. Russel, *J. Vet. Anim. Sci.* **25**, 681-686 (2001)
25. J. Achmadi, E. Pangestu, Surahmanto, A. Subrata, and M. A. Harahap, *IOP Conf. Ser. Earth Environ. Sci.* **518** (2020)